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NARCOTEST® Instructions for use

GENERAL GUIDELINES:

1. Test RESULTS are for PRESUMPTIVE IDENTIFICATION of illicit substances to help determine PROBABLE CAUSE only.
2. ALL PERSONNEL using this product should receive appropriate TRAINING.
3. TEST reagents may give FALSE POSITIVE or FALSE NEGATIVE results.
4. All test RESULTS should be confirmed by qualified scientists in the forensic lab.
5. SAMPLE CONTAMINATION from prior test can give MISLEADING results; always use a clean sampling device.
6. Test reagents #2, 3, 6, 7, 8, 14, 24, & 26 contain strong CORROSIVE acids and may cause bodily INJURY if spilled ... HANDLE WITH CARE.
7. Test reagents #5,7, 8, & 25 contain a FLAMMABLE substance and may ignite if exposed to flame... HANDLE WITH CARE.

FIRST AID PROCEDURES

INTERNAL:

CORROSIVE ACIDS: #2, 3, 6, 7, 8, 14, 24, & 26. DO NOT INDUCE VOMITING. Rinse mouth with water. Dilute swallowed acids with a glass of water.

ALKALI: Tests #9, 25, & 27... same as above

ALL OTHER TEST REAGENTS: #1, 4, 5, 13, & 23... INDUCE VOMITING.

EXTERNAL:

EYES: Holding the eyelid open, FLUSH GENTLY for 15 minutes with running water.

CLOTHING: Remove contaminated clothing and rinse skin thoroughly with water.

IF PAIN OR IRRITATION PERSISTS... SEEK COMPETENT MEDICAL ATTENTION PROMPTLY.

DESCRIPTION OF THE TEST UNIT

Individual test units consist of a flexible plastic tube in which a crushable glass ampoule(s) has been sealed. Most ampoules contain 0.5 ml of reagent. Reagents #1, 2, 3, 6, & 26 each have one ampoule. Reagents #4, 5, 7, 8, 9, 13, 14, 23, 24, 25, & 27 have a second ampoule in the cap. Care should be exercised in removing and replacing the cap to make sure the ampoule is not broken.

Apply pressure at the liquid level (or at the mid-point of the cap) to break the ampoules.

TESTING AN UNKNOWN MATERIAL

Testing an unknown is simple. It is important to thoroughly read the following procedure to familiarize oneself with all steps and operations. The following protocol applies to all materials tested. Refer to the **Individual Test Instructions** section for greater detail.

The correct test unit is opened by carefully removing the cap and the proper amount of suspect material (see next section) is placed in the tube. Hold the tube at the top between the thumb and forefinger to prevent static electricity from dispersing the sample ([see procedure in a pop-up window](#)). The cap is then replaced and the material forced to the bottom of the tube by tapping the tube on a hard surface. Each ampoule is broken by exerting sufficient pressure at the liquid level.

Always break the bottom ampoule first. DO NOT try to crush the broken glass. All that is needed is to release the chemical reagent sealed within the ampoule.

The reagents are mixed with the suspect material by flicking the side of the tube with the forefinger near the bottom after each ampoule is broken. The colors produced are compared with those on the proper chart immediately to no more than 60 seconds after breaking the final ampoule. The sample test unit is properly discarded after recording the color obtained from that particular test.

Note: INTENSE COLORS: May result from overloading or waiting too long to view the color, suggest repeating the test using less sample if required, or hold the tube almost horizontally to allow a thin layer of liquid to coat the wall of the tube. Light filtered through this thin layer will allow the color to be compared.

WEAK COLORS: This problem may occur with the hallucinogens or other weak samples. Repeat the test using two or three times the normal sample size.

QUANTITY OF SAMPLE NEEDED

Only a small quantity of suspect material is required to obtain a presumptive identification. A standard flat, wooden toothpick makes an inexpensive and easily obtained sampling device. Mark the broader end at a point 3/16 of an inch from the end. The suspect material can then be picked up to this point on the toothpick. This usually will provide between one (1) and three (3) milligrams for use as a sample. Fluffy drugs like cocaine HCl require a larger sample size. Familiarity with the testing procedure will eventually make it easier to judge the proper amount of sample needed. A pen knife or similar item may be used in place of the toothpick. Care should be taken to ensure that any measuring device is clean and free of contaminating material from previous tests.

PREPARATION OF SUSPECT MATERIAL

ALWAYS RETAIN SUFFICIENT SAMPLE OF SUSPECT MATERIAL FOR EVIDENTIAL ANALYSIS BY THE FORENSIC LABORATORY OR TOXICOLOGIST.

- Capsules.** Open the capsule, remove sufficient material for the test using a toothpick or other suitable device.
- Compressed.** Possibly hashish. Cut or scrape the block to provide small flakes for testing.
- blocks/pieces**
- Liquid.** Spread on a nonporous surface, allow to dry. Scrape to provide material for testing. Or... Absorb some on an uncolored, unscented tissue - allow to dry and use the tissue to test the unknown.
- Oils** A single drop of oil suspected of being hash oil is sufficient.
- Plant.** A pinch of dry, powdered plant material, or 8 - 10 flakes of fresh plant material is sufficient.
- material**
- Tablets.** Crush tablet to a fine powder between a fold of paper, remove sufficient material for the test using a toothpick or other suitable device.

A great deal of effort has gone into the preparation of the charts and color comparisons; however, colors described or printed are, at best, relative. Responses will be affected by purity and/or size of the sample, lighting conditions, temperature, how well the sample has been mixed with the reagent and a number of other factors. Only if the FULL TESTING SEQUENCE is followed, and common sense applied to reading color responses, will the results be conclusive.

HOW TO USE THE SEQUENTIAL TEST CHARTS

1. Follow the procedures outlined in this manual.
2. Run unknowns with Mayer's Reagent.
3. If a negative response (clear - no precipitate) is produced with Mayer's, go to CHART D.
4. If a positive response (white precipitate) is obtained with Mayer's, test the unknown with Marquis Reagent.
5. If a purple or violet color is produced with Marquis Reagent, use CHART A.
6. If a red, brown or orange color is produced with Marquis Reagent, use CHART B.
7. If no color is obtained with Marquis, use CHART C.
8. Once the proper chart has been selected, follow all of the test sequences indicated. Presumptive identification can be assumed if all color responses match those shown on the CHART.

DISPOSAL OF TEST UNITS

After color comparisons have been obtained and recorded, promptly dispose of the used test unit in a safe place. DO NOT STORE USED TEST UNITS! The contents of the used test may leak resulting in damage to clothing or bodily injury. If no safe disposal place is immediately available, place the used test units in one of the poly bags (part #7610) provided with the kit until such time as they can be safely discarded. DO NOT discard used #7603 units with used units of either #7607 or #7608.

- a Review: Do's and Don'ts -

Do...

1. Use care in removing and replacing caps.
2. Be sure measuring device is clean.
3. Use correct amount of suspect material.
4. Hold the test unit away from your face when loading, breaking ampoules, and agitating.
5. Apply pressure at the liquid level to break ampoules, and mix the reagent and sample thoroughly.
6. Compare the colors in a well-lighted area. Observe the colors formed immediately to NO MORE THAN ONE MINUTE after breaking the last ampoule.
7. Use a white background for viewing colors.
8. Dispose of used test units promptly and safely.
9. Store used test units in poly bag until disposed.
10. Flush all spilled reagents with water.
11. Treat burns from spilled reagents as any other burn.
12. Dispose of used #7603 test units promptly and in a separate poly bag. The nitric acid will partially dissolve the tube within 3-5 minutes, resulting in a leak of acid from the unit.

Don't...

1. Do Not crush the broken pieces of glass ampoule once broken. This will help

- avoid piercing the fingers and spilling reagent through a punctured tube.
2. Do Not shake the test units up and down to mix, or hold your face over the unit when loading or agitating.
 3. Do Not compare the colors in poor light such as under mercury or sodium vapor area lighting.
 4. Do Not hold the unit in front of colored surfaces for comparison.
 5. Do Not store used test units on person or in clothing.
 6. Do Not dispose of test units where accessible to children.
 7. Do Not ignore spilled reagents.
 8. Do Not dispose of #7603 with #7607 or #7608.
 9. Do Not place liquids directly into the tube.

INDIVIDUAL TEST INSTRUCTIONS

PROCEDURE A for 1 ampoule tests (1, 2, 3, 6, & 26):

1. Remove the cap.
2. Place sample in the tube.
3. Replace the cap.
4. Break the ampoule. Mix.
5. Observe color.

PROCEDURE B for 2 ampoule tests (4, 5, 7, 8, 9, 13, 14, 23, 24, 25, & 27):

1. Remove the cap.
2. Place sample in the tube.
3. Replace cap carefully.
4. Break bottom ampoule. Mix well.
5. Observe any color formed.
6. Break the cap ampoule and mix well.
7. Observe color.

Reagent 1 Mayer's Reagent (1 ampoule) A test for general narcotic compounds (this test is a prerequisite for all other tests, see the chart selector on the front side). (Use procedure A) The formation of a white to cream-colored gelatinous precipitate is indicative of the presence of one of the general narcotic compounds (or the amphetamines). Proceed to Test #2 (Marquis Reagent), a test for the opiates. If no precipitate is formed, proceed to Test #5 (Dille-Koppanyi Reagent), a test for barbiturates.

CONTENTS: 0.5 ml of a 1% potassium tri-iodo mercurate solution in water.

ANTIDOTE: Immediate dilution with water followed by INDUCED VOMITING using 2 tablespoons of syrup of ipecac. Seek medical advice.

Reagent 2 Marquis Reagent (1 ampoule) A test for opiates and amphetamine type compounds. (Use procedure A) Color responses: Violet to reddish-purple is indicative of the opiates. Proceed to Test #24 Mecke's for confirming heroin or #3 (nitric acid) to differentiate between heroin and morphine. An orange to red to brown sequence within 12 seconds may indicate the presence of an amphetamine. Brown may indicate demerol. Red may indicate the presence of mescaline.

CONTENTS: concentrated sulfuric acid with formaldehyde.

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 3 Nitric Acid (1 ampoule) This reagent is NOT a primary test. It is used to differentiate heroin from morphine and to confirm the identity of other suspect materials. (Use procedure A) Observe the color changes that take place. Colors: a. Yellow: heroin; b. Red to orange fading rapidly to yellow: morphine. CONTENTS: 0.5 ml of concentrated nitric acid. Nitric acid will dissolve the tube within 3-5 minutes. Place tube in a poly bag immediately after reading color. Do not dispose of #7603 with any other reagents.

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice. Caution: nitric acid is very reactive with human tissue, avoid contact.

Reagent 4 Cobalt Thiocyanate (2 ampoules) a test for cocaine HCl, procaine, dibucaine, and tetracaine. (Use procedure B) Observe the color formation after breaking the bottom ampoule. All of the above-named substances will produce an intense, brilliant blue, flaky precipitate. After breaking the cap ampoule: blue flakes remaining undissolved is indicative of cocaine. Blue flakes partially dissolved in the second solution is indicative of dibucaine. Blue flakes completely dissolved in the second solution is indicative of procaine or tetracaine.

CONTENTS: 0.5 ml stannous chloride, 5% aqueous (cap ampoule) and 0.5 ml cobalt thiocyanate 2% aqueous (bottom ampoule).

ANTIDOTE: Immediate dilution with water followed by INDUCED VOMITING using 2 tablespoons of syrup of ipecac. Seek medical advice.

Reagent 5 Dille-Koppanyl Reagent (2 ampoules) a test for barbiturates. (Use procedure B) A purple or reddish-violet color indicates the presence of a barbiturate. A pale, blue color is a negative response.

CONTENTS: 0.5 ml of 0.1% cobalt acetate in isopropanol (bottom ampoule) and 0.5 ml of 5% isopropanol-amine (cap ampoule).

ANTIDOTE: Immediate dilution with water followed by INDUCED VOMITING using 2 tablespoons of syrup of ipecac. Seek medical advice.

Reagent 6 Mandelin Reagent (1 ampoule) a confirming test for amphetamines and a presumptive test for Methadone. (Use procedure A) Observe the color changes. Greenish-brown changing to olive-green is indicative of the presence of DL- or D-amphetamines, such as benzedrine, dexedrine, obsedrin, etc. Olive-green changing to grey-green is indicative of methedrine (speed). Methadone turns an immediate deep blue. A brown color with a bluish tinge is indicative of an opiate.

CONTENTS: 0.5 ml of a 1% solution of ammonium vanadate in concentrated sulfuric acid.

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 7 Modified Ehrlich's Reagent (2 ampoules) a test for hallucinogens. Note: since the active ingredient in hallucinogens may be present in very small quantities, a larger sample may be necessary to produce a color response that can be compared easily. (Use procedure B) Colors: a. A slowly-developing (30-60 seconds) purple color is indicative of the presence of LSD or other ergot alkaloids. b. A wine pink color indicates the presence of DMT or DET.

CONTENTS: 0.5 ml concentrated hydrochloric acid (cap ampoule) and 0.5 ml of a 5% solution of p-dimethylamino benzaldehyde (bottom ampoule).

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 8 Duquenois Reagent (2 ampoules) a test for marihuana, hashish, THC and residues of THC in smoking paraphernalia. (Use procedure B) Agitate bottom ampoule one full minute before breaking the cap ampoule to allow the first reagent to work on the sample. Observe color development. A slowly developing grey-blue color or violet-blue color is indicative of marihuana or its active ingredient. NOTE: ALL PLANT MATERIAL and oils should be tested with Duquenois and if a positive result is obtained, confirmed by testing with KN Reagent #7609. Plant material failing to give a positive (blue) response to Duquenois or KN below should be examined by a forensic laboratory since many drugs can be sprayed on plant material other than marihuana (PCP on parsley for example).

CONTENTS: 0.5 ml of a 2% vanillin solution in alcohol (bottom ampoule) and 0.5 ml of concentrated hydrochloric acid (cap ampoule).

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting.

Reagent 9 KN Reagent (Fast Blue B Salt) (2 ampoules) a test for

marihuana, hashish, THC and residues of THC in smoking paraphernalia. (Use procedure B) Mix vigorously for at LEAST 30 seconds. Allow reagents to separate (layer). Observe the color in the bottom layer. Orange-red (reddish brown) to a very dark reddish brown is indicative of the presence of marihuana, hashish, THC and other cannabis products. ANY OTHER COLOR IN THE BOTTOM LAYER IS A NEGATIVE RESPONSE.

CONTENTS: 200 mgm % solution of Fast Blue B Salt in 0.5 ml of chlorinated hydrocarbon (bottom ampoule) and 0.5 ml 10% aqueous solution of sodium hydroxide (cap ampoule).

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 13 Cocaine Reagent - for Cocaine HCl and Cocaine Base (2 ampoules) (Use procedure B) Cocaine HCl and Cocaine Base will produce a flaky, intense blue precipitate in the bottom ampoule. Cocaine HCl and Base precipitates will not redissolve after breaking the cap ampoule.

CONTENTS: 0.5 ml 1% cobalt thiocyanate in 10% CH₃COOH (bottom ampoule); 0.5 ml stannous chloride 5% aqueous (cap ampoule).

ANTIDOTE: Immediate dilution with water followed by INDUCED VOMITING using 2 tablespoons of syrup of ipecac. Seek medical advice.

Reagent 14 Methaqualone, PCP Reagent (Use procedure B) This test is used to detect street level concentrations of PCP and as a test for methaqualone (quaalude). **Note:** other reagents in the sequential charts showing PCP reactions are for reasonably pure PCP, and will not react to "street" grades commonly found. Color should remain pink after breaking the bottom ampoule. Blue color forms with methaqualone or PCP after breaking the cap ampoule.

CONTENTS: 0.5 ml of 2% aqueous solution of cobalt thiocyanate (bottom ampoule); 0.2 ml 85% phosphoric acid (cap ampoule).

ANTIDOTE: DO NOT induce vomiting. Dilute with milk or water. Seek medical advice.

Reagent 23 Sodium Nitroprusside for Methamphetamine (2 ampoules) (Use procedure B) Place a **very** small amount of suspect material into the tube.

An immediate dark blue color indicates the presence of methamphetamine.

Note: a similar reaction may occur with "XTC" (MDMA). Distinguish in Marquis Reagent #2: "XTC" will form a purple/black color while meth is a rapid orange, to red, to brown within 12 seconds. A negative test (no meth present) is salmon colored.

CONTENTS: Water, Sodium Nitroprusside, Sodium Carbonate.

ANTIDOTE: If swallowed, INDUCE VOMITING. Seek medical advice.

Reagent 24 Mecke's (Modified) Reagent A test for Heroin. (2 ampoules)

(Use procedure B) Agitate bottom ampoule for 30 seconds disregarding any color generated at this point.* Break cap ampoule and agitate for 5 seconds.

Solution turns immediate green in the presence of heroin.

CAUTION! Contains Corrosive Acid (sulfuric acid in both ampoules). Keep out of the reach of children. Do Not Store after breaking ampoules. Do not hold close to the face when breaking ampoules or agitating. Promptly dispose.

ANTIDOTE: Immediate dilution with water. DO NOT induce vomiting. Seek medical advice.

*Note sole exception: A slowly developing purple (4-5 seconds) in the first ampoule may indicate MDMA ("XTC") going to a deep brown in the second ampoule.

Reagent 25 Valium®/Diazepam Clonazepam/Rohypnol® "roofies" (2 ampoules) (Use procedure B) Solution turns from pale violet to purple within sixty seconds.

CONTENTS: 0.2 ml 2M KOH in methanol (bottom ampoule) and 0.2 ml 0.2% m-dinitrobenzene in isopropanol (cap ampoule).

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 26 Talwin® (Pentazocine) (Fröhdes reagent) (1 ampoule) (Use procedure A) Solution turns a bright blue instantly. Talwin is the registered trade name of Winthrop Laboratories. Confirm in Test #6, Talwin turns olive green.

CONTENTS: 0.5 ml 0.5% ammonium molybdate in H₂SO₄.

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 27 Ephedrine (Modified Chen's) Reagent (2 ampoules) (Use procedure B) No color is formed after the bottom ampoule is broken. Solution turns bright blue instantly on breaking the cap ampoule. Note: the reagent itself when mixed is aqua-blue green.

CONTENTS: Bottom ampoule 0.5 ml 1% Copper sulfate in 1% CH₃COOH. Cap ampoule 0.2 ml 2N NaOH.

ANTIDOTE: Immediate dilution with water is recommended. DO NOT induce vomiting. Seek medical advice.

Reagent 28 GHB Reagent (1 ampoule) (Use procedure B) If sample is liquid use two to five drops with supplied disposable pipette. Solution turns green to blue-green in the presence of GHB. Do not store tube after breaking ampoule. Dispose of the pipette properly after use (never reuse).

ANTIDOTE: Wash out mouth with water and INDUCE VOMITING using two tablespoonsful of syrup of ipecac. Seek medical advice.