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# **Opioids and efflux transporters. Part 3: P-glycoprotein** substrate activity of 3-hydroxyl addition to meperidine analogs

Susan L. Mercer, Christopher W. Cunningham, Natalie D. Eddington and Andrew Coop\*

Department of Pharmaceutical Sciences, University of Maryland School of Pharmacy, 20 Penn Street, Room 543, Baltimore, MD 21201, USA

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Abstract—Numerous studies have shown that many clinically employed opioid analgesics are substrates for P-glycoprotein (P-gp), suggesting that up-regulation of P-gp may contribute to the development of central tolerance to opioids. The studies herein focus on the development of SAR for P-gp substrate activity in the meperidine series of opioids. Addition of a 3-OH to meperidine and the ketone analog of meperidine yielding bemidone and ketobemidone, respectively, significantly increased P-gp substrate affinity. The results of this study have implications in the development of novel analgesics to be utilized as tools to study the contribution of P-gp on the development of central tolerance to opioids. © 2008 Elsevier Ltd. All rights reserved.

*Introduction.* There is a growing body of evidence that suggests efflux transporters, specifically P-glycoprotein (P-gp), may play a role in the development of opioid related central tolerance and constipation.<sup>1-6</sup> Recent studies have shown that opioids are substrates for P-gp, although to differing extents,<sup>7</sup> and P-gp is up-regulated at the blood-brain barrier (BBB) of morphine<sup>3</sup> (1) and oxycodone<sup>2</sup> (2) (Fig. 1) tolerant rats. Upon chronic administration, the up-regulated P-gp would be expected to result in lower brain concentrations of opioid, thereby exacerbating tolerance to the central analgesic effects. P-gp knockout animals<sup>8</sup> are available and offer a useful model to study the effects of P-gp on opioids; however, an alternative approach in wild-type animals is the development of mu opioid receptor agonists which are not P-gp substrates. These compounds would allow a full investigation of the contribution of up-regulated P-gp to opioid tolerance, as full cross-tolerance between morphine and the opioid lacking P-gp substrate activity would not be anticipated to occur. Additionally, opioids lacking P-gp substrate activity may potentially be developed into analgesics with lower degrees of tolerance.

Meperidine (3), a moderately potent mu opioid analgesic, <sup>9,10</sup> has been reported to possess low P-gp substrate activity.<sup>7</sup> Therefore, our investigations are focused on delineating the structure-activity relationship (SAR) for the addition of a *m*-OH, while increasing mu opioid potency based on known SAR for this series.<sup>10</sup>

*Results and discussion.* The compounds synthesized are readily known in the literature as mu opioid analgesics;<sup>10</sup> however, the syntheses described here are novel approaches. Meperidine (**3**) was prepared from nitrile **4** (obtained from Sigma–Aldrich, Inc.), via alkylation with MeI in DMF in the presence of  $K_2CO_3$ , followed by aqueous NH<sub>4</sub>Cl hydrolysis of the *N*-methyl nitrile **5** to the ethyl ester through treatment with H<sub>2</sub>SO<sub>4</sub> and EtOH. Treatment of **5** with EtMgBr, via a Grignard reaction,<sup>11</sup> produced the ketone meperidine analog **6** (Scheme 1).

Bemidone 9 was prepared from the condensation of mechloroethamine hydrochloride and 3-methoxyphenyl-



Figure 1. Morphine (1) and oxycodone (2).

Keywords: Meperidine; P-glycoprotein; Opioids; Tolerance.

<sup>\*</sup> Corresponding author. Tel.: +1 410 706 2029; fax: +1 410 706 5017; e-mail: acoop@rx.umaryland.edu

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Scheme 1. Reagents and conditions: (a) MeI,  $K_2CO_3$ , DMF; (b)  $H_2SO_4$ , EtOH, reflux; (c) EtMgBr, NH<sub>4</sub>Cl hydrolysis.

acetonitrile (both reagents obtained from Sigma–Aldrich, Inc.) with NaH and NaOH to yield 7. *O*-dealkylation of 7 was performed with BBr<sub>3</sub> and NH<sub>4</sub>OH,<sup>12</sup> converting the methoxy group to a phenol **8**, followed by nitrile hydrolysis to give the *m*-OH ethyl ester as previously described.<sup>4</sup> Treatment of 7 with an EtMgBr Grignard reagent,<sup>11</sup> followed by aqueous NH<sub>4</sub>Cl hydrolysis produced **10**, which then underwent treatment with BBr<sub>3</sub> to produce ketobemidone (**11**) (Scheme 2).

All compounds were converted to their respective salts (see Table 1). Drug stimulated P-gp ATPase activity

was estimated using the Pgp-Glo assay system<sup>13</sup> (Promega, Madison, WI) and the results are shown in Table 1. Briefly, this method relies on the ATP dependence of the light-generating reaction of firefly luciferase where ATP consumption is detected as a decrease in luminescence, the greater the decrease in signal the higher the P-gp activity. Sodium orthovanadate was used as a P-gp ATPase inhibitor, whereas verapamil was used as a positive control. All test compounds were analyzed at 200  $\mu$ M and fold stimulation values were calculated using Eq. 1. Fold stimulation values greater than 2.0 indicate a P-gp substrate.<sup>14</sup>

## Fold stimulation by a test compound

$$=\frac{\text{Test compound stimulated P-gp activity}}{\text{Basal P-gp activity}}$$
(1)

The addition of a *m*-OH into the phenyl ring significantly increases the P-gp fold stimulation of meperidine analogs. Meperidine itself has a P-gp fold stimulation value of 1.78 and increases to 2.64 with the *m*-OH addition (bemidone, 9). Whereas the ketone analog 6, with a P-gp fold stimulation value of 1.37, increases to 4.89 with the *m*-OH addition (ketobemidone, 11). Thus, the addition of a *m*-OH increases the P-gp substrate activity of these meperidine analogs, which are members of the 4-phenylpiperidine class of opioids.

The hydroxylated meperidine analogs were initially pursued to investigate the relationship between P-gp and increased opioid potency. Interestingly, these results are consistent with previous studies in our laboratory which



Scheme 2. Reagents: (a) NaH, NaOH; (b) BBr<sub>3</sub>, NH<sub>4</sub>OH; (c) H<sub>2</sub>SO<sub>4</sub>, EtOH; (d) EtMgBr, NH<sub>4</sub>Cl hydrolysis.

Table 1. Fold stimulation values of test compounds prepared, salt form, yield, and melting point

Compound	Name	Salt	Yield (%)	Mp (°C)	Fold stimulation ± SEM
	Non-treated (control)				1.00
3	Meperidine	Oxalate	7	190-192	$1.78 \pm 0.39^*$
6	Ketone analog	Citrate	56	170-171	$1.37 \pm 0.19^*$
9	Bemidone	Oxalate	36	200-202	$2.64 \pm 0.82^{*}$
11	Ketobemidone	Oxalate	51	233–235	$4.89 \pm 1.94^*$

All compounds assayed at 200  $\mu$ M. Data are represented as fold stimulation ± SEM (n = 3). \*Significant difference (p < 0.05) from control (non-treated) as determined from *t*-test. All compounds gave satisfactory CHN (±0.4%) and spectral analysis (see Supporting Information).

showed that removal of the 3- and 6-OH from morphine resulted in decreased P-gp substrate activity,<sup>5</sup> as morphine is a P-gp substrate.<sup>7</sup> These studies attest that the *m*-OH substituent increases P-gp substrate activity across the phenylpiperidine and morphinan classes of opioids. Furthermore, the development of opioids lacking P-gp substrate activity should not possess a *m*-OH substituent. The interaction between opioids and P-gp is currently under investigation and these results aid in further SAR development. The ultimate goal is to develop a potent opioid with low P-gp substrate activity for use as a tool to study the contribution of P-gp up-regulation to the development of opioid tolerance and crosstolerance between opioids with P-gp substrate activity and those without.

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# Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2008. 04.046.

## **References and notes**

- Aquilante, C. L.; Letrent, S. P.; Pollack, G. M.; Brouwer, K. L.. Life Sci. 2000, 66, PL47.
- Hassan, H. E.; Myers, A. L.; Lee, I. J.; Coop, A.; Eddington, N. D. J. Pharm. Sci. 2007, 96, 2494.
- 3. Ambudkar, S. V.; Kimchi-Sarfaty, C.; Sauna, Z. E.; Gottesman, M. M. Oncogene 2003, 22, 7468.
- 4. Mercer, S. L.; Hassan, H. E.; Cunningham, C. W.; Eddington, N. D.; Coop, A. *Bioorg. Med. Chem. Lett.* 2007, 17, 1160.
- Cunningham, C. W.; Mercer, S. L.; Hassan, H. E.; Traynor, J. R.; Eddington, N. D.; Coop, A. J. Med. Chem. 2008, 51, 2316.
- Hassan, H. E.; Mercer, S. L.; Cunningham, C. W.; Coop, A.; Eddington, N. D. Int. J. Pharm. Sci. 2008, accepted for publication.
- 7. Dagenais, C.; Graff, C. L.; Pollack, G. M. Biochem. Pharmacol. 2004, 67, 269.
- Schinkel, A. H.; Smit, J. J.; van Tellingen, O.; Beijnen, J. H.; Wagenaar, E.; van Deemter, L.; Mol, C. A.; van der Valk, M. A.; Robanus-Maandag, E. C.; te Riele, H. P. *Cell* 1994, 77, 491.
- 9. Janssen, P. A.; Eddy, N. B. J. Med. Pharm. Chem. 1960, 2, 31.
- 10. Casy, A. F.; Parfitt, R. T. *Opioid Analgesics: Chemistry and Receptors*; Plenum Press: New York, 1986, (Chapter 6).
- 11. Kharasch, M. S.; Reinmuth, O. *Grignard Reactions on Nonmetallic Substances*; Prentice Hall: New York, 1954, (Chapter 10).
- 12. Rice, K. C. J. Med. Chem. 1977, 20, 164.
- Ambudkar, S. V.; Dey, S.; Hrycyna, C. A.; Ramachandra, M.; Pastan, I.; Gottesman, M. M. Annu. Rev. Pharmacol. Toxicol. 1999, 39, 361.
- Polli, J. W.; Wring, S. A.; Humphreys, J. E.; Huang, L.; Morgan, J. B.; Webster, L. O.; Serabjit-Singh, C. S. J. Pharmacol. Exp. Ther. 2001, 299, 620.