## Review of Chemical Syntheses of 7-Keto- $\Delta^5$ -sterols

Edward J. Parish\*, Stephen A. Kizito, and Zhihai Qiu

Department of Chemistry, Auburn University, Auburn, Alabama 36849

**ABSTRACT:** Steroids bearing ketone functionality at carbon-7 are found commonly in nature, and the most prevalent of these are the 7-keto- $\Delta^5$ -sterols. These substances have diverse biological properties and are present in biological samples and food products. For the purpose of studying this class of oxysterols, many chemical methods, involving the chemical oxidation of  $\Delta^5$ -sterols to the corresponding 7-keto- $\Delta^5$ -sterol derivatives have been developed to produce these compounds. We have undertaken a review and evaluation of chemical methods for the synthesis of these compounds and have endeavored to enhance one of these procedures to yield products for chemical and biological investigations.

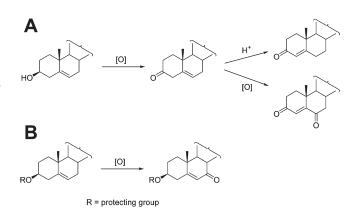
Paper no. L9537 in Lipids 39, 801-804 (August 2004).

Some of the most frequently encountered oxysterols are those with a ketone function at carbon-7. The major portion of these is the 7-keto- $\Delta^5$ -sterols, which originate from the oxidation of  $\Delta^5$ -sterols. These compounds, which are found in animal tissues, food products (1,2), and certain folk medicines (3–6), are significant inhibitors of HMG-CoA reductase (7,8), sterol synthesis (7,9), and cell replication (10–12).

The chemical synthesis of 7-keto- $\Delta^5$ -sterols relies on the allylic oxidation of carbon-7 of  $\Delta^5$ -sterols. We have studied a number of different synthetic methods and procedures and review our findings here.

In many chemical and biological studies, 7-keto- $\Delta^5$ -sterols containing the 3 $\beta$ -hydroxyl group are the desired compounds. During the chemical (allylic) oxidation of the corresponding  $\Delta^5$ sterols, the 3 $\beta$ -hydroxyl group must be protected, usually as the acetate or benzoate ester (13,14), to avoid oxidation of the C-3 hydroxyl group to a ketone. The benzoate derivative is usually preferred owing to its superior crystallinity (i.e., ease of crystal formation leading to higher yields) when purified by recrystallization. Figure 1A demonstrates the reaction paths of oxidation that occur when the 3 $\beta$ -hydroxyl is not protected (15–19).

The most common methods of allylic oxidation of  $\Delta^5$ sterols are those that rely on the use of chromium(VI) reagents. Early or "classical" methods of oxidation used chromium trioxide (CrO<sub>3</sub>) (20–23) or sodium chromate (24–26) and *t*-butyl chromate (27,28) in acetic acid; these afforded only limited success and produced the 7-keto compounds in modest yields. For example, the allylic oxidation of cholesteryl benzoate with sodium chromate in acetic acid/acetic anhydride gave 7-ketocholesteryl benzoate in an



**FIG. 1.** Chemical oxidation of  $3\beta$ -hydroxy- $\Delta^5$ -sterols (A) in the absence of a protection for the  $\beta$ -hydroxyl group and (B) in the presence of a protective group.

optimal yield of 38% (29). Figure 1B demonstrates the reaction that occurs when the  $3\beta$ -hydroxyl group is protected.

Synthetically useful changes in the properties and reactivity of chromium(VI) reagents have been brought about by the formation of amine complexes. The Collins reagent is formed by the complexation of chromium trioxide with pyridine (30,31). With this reagent, the allylic oxidation of cholesteryl benzoate gave a 68% yield of 7-ketocholesteryl benzoate (32), and in a related study, using anhydrous conditions, cholesteryl acetate was oxidized to 7-ketocholesteryl acetate in 72% yield (33). Similar complexes have been formed using chromium trioxide and pyrazole (34), 3,5-dimethylpyrazole (29), and benzotriazole (35) and have been shown to oxidize cholesteryl benzoate to 7-ketocholesteryl benzoate in 70–76% yields. These reactions require the preparation of the reagent complex before each reaction.

Commercially available pyridinium chlorochromate (PCC) has been widely used in organic synthesis for the oxidation of primary and secondary alcohols to carbonyl compounds (36). This reagent, in methylene chloride containing pyridine (37), other aromatic amines (38), pyrazole (39), 3,5-dimethylpyrazole (40), and benzotriazole (35), was reported to effect the selective oxidation of the allylic hydroxyl function of a number of steroidal alcohols. At room temperature, PCC in methylene chloride was an ineffective reagent for allylic oxidation (29). In contrast to these results, we have achieved moderate success by using PCC in refluxing methylene chloride for allylic and benzylic oxidations (41). PCC in DMSO also has been used for the oxidation of  $\beta$ -ionone to the corresponding diketone (42).

<sup>\*</sup>To whom correspondence should be addressed. E-mail: parisej@auburn.edu Abbreviations: PCC, pyridinium chlorochromate; PFC, pyridinium fluorochromate; TBHP, *t*-butyl hydroperoxide.

In a related study, we found that benzene was a superior solvent for allylic oxidations using PCC (43). PCC in refluxing benzene could effect a high-yield (87%) oxidation of cholesteryl benzoate to 7-ketocholesteryl benzoate. This conversion was accomplished with a 1:30 ratio of reagent (PCC) when 1–10 g of cholesteryl benzoate was oxidized. Oxidation of quantities of less than 1 g cholesteryl benzoate was successfully performed by using smaller quantities of reagent (1:25) with similar yields, thus demonstrating the usefulness of the described method for both large- and small-scale preparations. This efficient (i.e., high-yielding) procedure represents a significant improvement in both yield and convenience compared with other reported methods for the allylic oxidation of cholesteryl benzoate to 7-ketocholesteryl benzoate.

With the reaction conditions described (43), we conducted additional studies using other solvents with cholesteryl benzoate and PCC. Under these conditions, refluxing acetone, pyridine, *N*,*N*-dimethylformamide, and DMSO at 100°C yielded 7-ketocholesteryl benzoate in 2, 0, 18, and 77% yield, respectively. In an earlier study, we found that using refluxing methylene chloride as solvent yielded 54% of 7-ketocholesteryl benzoate from cholesteryl benzoate (41).

In another additional study, we showed that pyridinium fluorochromate (PFC), in refluxing benzene, was also an effective and convenient reagent for the efficient allylic oxidation of  $\Delta^5$ -sterols to the corresponding C-7 unsaturated ketones in high yields (44). With this reagent, the allylic oxidation of cholesteryl benzoate resulted in an 88% yield of 7-ketocholesteryl benzoate. In the same study, cholesteryl acetate was oxidized to its 7-keto derivative in an 87% yield.

Other oxidation studies have used hydroperoxides [e.g., t-butyl hydroperoxide (TBHP)] with different types of catalysts. Chromium trioxide (45) and bis(tributyltin oxide)dioxochromium (46) have been used as catalysts to obtain 7-keto- $\Delta^5$ -sterols. However, epoxidation of the double bond was also observed. Good yields of these products were also reported when the reaction with the oxidizing agent TBHP was catalyzed by hexacarbonyl chromium (47,48) and ruthenium trichloride (49). However, the high toxicity of hexacarbonyl chromium, the high cost of the ruthenium catalyst, and potential safety risks have led researchers to explore other methods (50). The use of TBHP with catalysts such as Cu(I), Cu(II), or Cu metal (51) gave good yields in the allylic oxidation of  $\Delta^5$ -sterols and required small amounts of reagents and solvents, and the copper catalysts were inexpensive and less toxic than the chromium reagents.

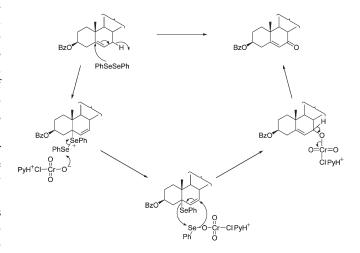
More environmentally friendly oxidations by molecular oxygen and *N*-hydroxyphthalimide as catalyst (52,53) give good yields of 7-keto- $\Delta^5$ -sterols. These methods are readily applicable and inexpensive, and the catalyst can be recovered. However, the required use of an oxygen atmosphere makes them inconvenient (50). In a related procedure, allylic oxidation at the C-7 position was accomplished using *N*-hydroxyphthalimide-catalyzed oxidation in air with benzoyl peroxide as a free radical initiator (54,55). The resulting C-7 hydroperoxide was dehydrated with copper(II) chloride in pyridine to produce the corresponding C-7 ketone of stigmasterol in 81-82% yield. In addition to being environmentally friendly, this procedure had an added advantage in that no protecting group was required for the 3 $\beta$ -hydroxyl group, since it was not oxidized under these reaction conditions.

Recently, TBHP has been used in the presence of copper(I) iodide, and tetra-*n*-butylammonium bromide was used as a phase-transfer catalyst in a two-phase system of water and methylene chloride (56,57). The allylic oxidation was found to proceed more efficiently when TBHP was added to the reaction mixture in portions. The high-yield conversion (>70%) of  $\Delta^5$ -sterols into the corresponding C-7 unsaturated ketones in short reaction times was reported.

We have continued our studies on the allylic oxidation of  $\Delta^5$ -sterols using PCC to produce the C-7 ketones. The use of both PCC and PFC in refluxing benzene represents one of the most convenient methods for the allylic oxidation of these substrates (43,44,58). However, long reaction times, large volumes of solvent, and high molar ratios of the oxidant are required.

To improve on these procedures, we tested the use of diphenyl diselenide as a catalyst to reduce reaction times and amounts of reagent required for oxidation and observed that lower quantities of the required reagent (PCC) were needed. That selenium reagents introduce a hydroxyl group at allylic positions in a substrate has long been known (59–61). In our case these substrates would be oxidized to a ketone. The involvement of the catalyst in this reaction would alter the mechanism and would be similar to that already described for the enhanced allylic chlorination of  $\beta$ -pinene using diphenyl diselenide as a catalyst (62).

When we modified our established procedure (43), PCC (1:5 molar ratio of steroid substrate/PCC) and catalytic amounts of diphenyl diselenide (1:0.1 molar ratio of steroid substrate/diphenyl diselenide) were heated together in refluxing benzene with cholesteryl benzoate; allylic oxidation at the



**FIG. 2.** Mechanism of allylic oxidation of cholesteryl benzoate by pyridinium chlorochromate and a diphenyl diselenide catalyst.

C-7 position occurred in 5 h, and 7-ketocholesteryl benzoate was obtained in 50% yield. Continued heating for a total of 24 h produced a 76% yield. When this reaction was conducted in the absence of a diphenyl diselenide catalyst, an 11% yield of the C-7 ketone was obtained after 24 h of reaction. Without the catalyst, a molar ratio of 1:30 of PCC was required for optimal product yield (87%). A proposed mechanism for the allylic oxidation of cholesteryl benzoate using PCC and diphenyl diselenide is shown in Figure 2. The ionic mechanism shown is supported by the addition of 2,2'-azobisisobutyronitrile, a radical initiator (63), which produced no observed enhancement of the reaction rate (monitored by TLC) or final yield of the reaction product.

In conclusion, we believe the results presented herein provide useful information concerning the chemical synthesis of 7-keto- $\Delta^5$ -sterols resulting from the allylic oxidation of  $\Delta^5$ sterols. The major methods of synthesis have been reviewed and the advantages of certain procedures have been indicated. In addition, we have attempted to develop novel and efficient approaches to the synthesis of these products.

## REFERENCES

- 1. Smith, L.L. (1981) *Cholesterol Autoxidation*, Plenum Press, New York.
- Guardiola, F., Dutta, P.C., Codony, R., and Savage, G.P. (eds.) (2002) Cholesterol and Phytosterol Oxidation Products: Analysis, Occurrence, and Biological Effects, AOCS Press, Champaign, 394 pp.
- Hietter, H., Bischoff, P., Beck, J.P., Ourisson, G., and Luu, B. (1986) Comparative Effects of 7β-Hydroxycholesterol Towards Murine Lymphomas, Lymphoblasts and Lymphocytes: Selective Cytotoxicity and Blastogenesis Inhibition, *Cancer Biochem. Biophys.* 9, 75–83.
- Cheng, K.P., Nagano, H., Luu, B., Ourisson, G., and Beck, J.P. (1977) Chemistry and Biochemistry of Chinese Drugs. Part I. Sterol Derivatives Cytotoxic to Hepatoma Cells, Isolated from the Drug Bombyx cum Botryte, *J. Chem. Res.* (S), 217; (M), 2501–2521.
- Nagano, H., Poyser, J.P., Cheng, K.-P., Luu, B., Ourisson, G., and Beck, J.P. (1977) Chemistry and Biochemistry of Chinese Drugs. Part II. Hydroxylated Sterols, Cytotoxic Towards Cancerous Cells: Synthesis and Testing, *J. Chem. Res.* (S), 218; (M), 2522–2571.
- Zander, M., Koch, P., Luu, B., Ourisson, G., and Beck, J.P. (1977) Chemistry and Biochemistry of Chinese Drugs. Part III. Mechanism of Action of Hydroxylated Sterols on Cultured Hepatoma Cells, J. Chem. Res. (S), 219; (M), 2572–2584.
- Kandutsch, A.A., Chen, H.W., and Heiniger, H.J. (1978) Biological Activity of Some Oxygenated Sterols, *Science 201*, 498–501.
- Taylor, F.R., Saucier, S.E., Shown, E.P., Parish, E.J., and Kandutsch, A.A. (1984) Correlation Between Oxysterol Binding to a Cytosolic Binding Protein and Potency in the Repression of Hydroxymethylglutaryl Coenzyme A Reductase, *J. Biol. Chem.* 259, 12382–12387.
- Schroepfer, G.J., Jr. (2000) Oxysterols: Modulators of Cholesterol Metabolism and Other Processes, *Physiol. Rev.* 80, 361–554.
- Schroepfer, G.J., Jr. (1981) Sterol Biosynthesis, Annu. Rev. Biochem. 50, 585–621.
- 11. Parish, E.J., Chitrakorn, S., Luu, B., Schmidt, G., and Ourisson,

G. (1989) Studies of the Oxysterol Inhibition of Tumor Cell Growth, *Steroids* 53, 579–596.

- Guardiola, F., Codony, R., Addis, P.B., Rafecas, M., and Boatella, J. (1996) Biological Effects of Oxysterols: Current Status, *Food Chem. Toxicol.* 34, 193–211.
- Kumar, V., Amann, A., Ourisson, G., and Luu, B. (1987) Stereospecific Syntheses of 7α- and 7β-Hydroxycholesterols, *Synth. Commun.* 17, 1279–1286.
- Atwater, N.W. (1961) Oxasteroids. II. 6-Oxaandrostane Derivatives, J. Am. Chem. Soc. 83, 3071–3079.
- Dhar, D.N., and Singh, A.K. (1977) Pyridinium Chlorochromate Oxidation of Some Steroidal Systems. Regioselective Opening of Ring 'F' in Spirostan, Z. Naturforsch. B: Anorg Chem., Org. Chem. 32B, 1476–1477.
- Parish, E.J., Chitrakorn, S., Taylor, F.R., and Saucier, S.E. (1984) Chemical Synthesis of 4,4'-Dimethyl-7-oxygenated Sterols. Inhibitors of 3-Hydroxy-3-methylglutaryl Reductase, *Chem. Phys. Lipids* 36, 179–188.
- 17. Parish, E.J., and Honda, H. (1990) A Facile Synthesis of Steroidal  $\Delta^4$ -3-Ketones Using Pyridinium Chlorochromate (PCC), *Synth. Commun.* 20, 1167–1174.
- Parish, E.J., Honda, H., Chitrakorn, S., and Livant, P. (1991) A Facile Chemical Synthesis of Cholest-4-en-3-one. Carbon-13 Nuclear Magnetic Resonance Spectral Properties of Cholest-4en-3-one and Cholest-5-en-3-one, *Lipids* 26, 675–677.
- 19. Parish, E.J., Kizito, S.A., and Heidepriem, R.W. (1993) A Novel Synthesis of Steroidal  $\Delta^4$ -3,6-Diones Using Pyridinium Chlorochromate (PCC), *Synth. Commun.* 23, 223–230.
- Marker, R.E., Kamm, O., Fleming, G.H., Popkin, A.H., and Wittle, E.L. (1937) Sterols. X. Cholesterol Derivatives, J. Am. Chem. Soc. 59, 619–621.
- Stavely, H.E., and Bollenback, G.N. (1943) Steroids with Double Bonds Between Quaternary Carbon Atoms. I. The Oxidation of α-Ergostenyl Acetate, *J. Am. Chem. Soc.* 65, 1285–1289.
- Klyne, W. (1951) Some 7-Substituted Derivatives of the 5α-Pregnane Series, J. Chem. Soc., 3449–3451.
- Kasal, A. (2000) Epalons: 6-Substituted Derivatives of 7-Norepiallopregnanolone, *Tetrahedron* 56, 3559–3565.
- Marshall, C.W., Ray, R.E., Laos, I., and Riegel, B. (1957) 7-Oxo Steroids. II. Steroidal 3β-Hydroxy-Δ<sup>5</sup>-7-ones and -Δ<sup>3,5</sup>-7-ones, *J. Am. Chem. Soc.* 79, 6308–6313.
- Amann, A., Ourisson, G., and Luu, B. (1987) Stereospecific Syntheses of the Four Epimers of 7,22-Dihydroxycholesterol, *Synthesis*, 1002–1005.
- Cook, R.P. (1958) Cholesterol: Chemistry, Biochemistry, and Pathology, pp. 56–57, Academic Press, New York.
- Marshall, C.W., Ray, R.E., Laos, I., and Riegel, B. (1957) 7-Oxo Steroids. I. Steroidal 3-Hydroxy-3,5-dien-7-ones, *J. Am. Chem. Soc.* 79, 6303–6308.
- Singh, H., Bhardwaj, T.R., and Paul, D. (1977) Steroids and Related Studies. Part 41. Schmidt Reaction with 3β-Acetoxypregn-5-ene-7,20-dione, *J. Chem. Soc., Perkin Trans. I*, 1987–1989.
- Salmond, W.G., Barta, M.A., and Havens, J.L. (1978) Allylic Oxidation with 3,5-Dimethylpyrazole-Chromium Trioxide Complex. Steroidal Δ<sup>5</sup>-7-Ketones, J. Org. Chem. 43, 2057–2059.
- Collins, J.C., Hess, W.W., and Frank, F.J. (1968) Dipyridine-Chromium(VI) Oxide Oxidation of Alcohols in Dichloromethane, *Tetrahedron Lett.*, 3363–3366.
- Dauben, W.G., Lorber, M.E., and Fullerton, D.S. (1969) Allylic Oxidation of Olefins with Chromium Trioxide Pyridine Complex, *J. Org. Chem.* 34, 3587–3592.
- 32. Cook, R.P. (1958) Cholesterol: Chemistry, Biochemistry, and Pathology, p. 100, Academic Press, New York.
- Fullerton, D.S., and Chen, C.-M. (1976) *In situ* Allylic Oxidations with Collins Reagent, *Synth. Commun.* 6, 217–220.
- 34. Parish, E.J., Chitrakorn, S., and Todd, K.L., III (1985) Steroidal

Allylic Oxidation with Chromium Trioxide in the Presence of Pyrazole, *Org. Prep. Proced. Int.* 17, 192–194.

- Parish, E.J., and Chitrakorn, S. (1985) Benzotriazole-Mediated Selective Chromium(VI) Oxidations, *Synth. Commun.* 15, 393–399.
- Piancatelli, G., Scettri, A., and D'Auria, M. (1982) Pyridinium Chlorochromate: A Versatile Oxidant in Organic Synthesis, *Synthesis*, 245–258.
- Parish, E.J., and Schroepfer, G.J., Jr. (1980) Selective Oxidation of Steroidal Allylic Alcohols, *Chem. Phys. Lipids* 27, 281–288.
- Parish, E.J., Scott, A.D., Dickerson, J.R., and Dykes, W. (1984) Further Studies on the Selective Oxidation of Steroidal Allylic Alcohols, *Chem. Phys. Lipids* 35, 315–320.
- Parish, E.J., Chitrakorn, S., and Lowery, S. (1984) Selective Oxidation of Steroidal Allylic Alcohols Using Pyrazole and Pyridinium Chlorochromate, *Lipids 19*, 550–552.
- Parish, E.J., and Scott, A.D. (1983) Selective Oxidation of Steroidal Allylic Alcohols Using 3,5-Dimethylpyrazole and Pyridinium Chlorochromate, J. Org. Chem. 48, 4766–4768.
- Parish, E.J., Chitrakorn, S., and Wei, T.Y. (1986) Pyridinium Chlorochromate-Mediated Allylic and Benzylic Oxidation, *Synth. Commun.* 16, 1371–1375.
- Becher, E., Albrecht, R., Bernhard, K., Leuenberger, H.G.W., Mayer, H., Mueller, R.K., Schuep, W., and Wagner, H.P. (1981) Synthesis of Astaxanthin from β-Ionone. I. Enantiomeric C<sub>15</sub>-Wittig Salts by Chemical and Microbial rResolution of (±)-3-Acetoxy-4-oxo-β-ionone, *Helv. Chim. Acta* 64, 2419–2435.
- Parish, E.J., Wei, T.-Y., and Livant, P. (1987) A Facile Synthesis and Carbon-13 Nuclear Magnetic Resonance Spectral Properties of 7-Ketocholesteryl Benzoate, *Lipids* 22, 760–763.
- 44. Parish, E.J., Sun, H., and Kizito, S.A. (1996) Allylic Oxidation of  $\Delta^5$ -Steroids with Pyridinium Fluorochromate, *J. Chem. Res.* (*S*), 544–545.
- 45. Muzart, J. (1987) Synthesis of Unsaturated Carbonyl Compounds via a Chromium-Mediated Allylic Oxidation by 70% tert-Butyl Hydroperoxide, Tetrahedron Lett. 28, 4665-4668.
- 46. Muzart, J. (1989) Bimetallic Oxidation Catalysts: Oxidations with *tert*-Butyl Hydroperoxide Mediated by Bis(tributyltin oxide)dioxochromium(VI), *Synth. Commun.* 19, 2061–2067.
- 47. Pearson, A.J., Chen, Y.S., Hsu, S.Y., and Ray, T. (1984) Oxidation of Alkenes to Enones Using *tert*-Butyl Hydroperoxide in the Presence of Chromium Carbonyl Catalysts, *Tetrahedron Lett.* 25, 1235–1238.
- 48. Pearson, A.J., Chen, Y.S., Han, G.R., Hsu, S.Y., and Ray, T. (1985) A New Method for the Oxidation of Alkenes to Enones. An Efficient Synthesis of  $\Delta^5$ -7-Oxo Steroids, J. Chem. Soc., Perkin Trans. I, 267–273.
- Miller, R.A., Li, W., and Humphrey, G.R. (1996) A Ruthenium-Catalyzed Oxidation of Steroidal Alkenes to Enones, *Tetrahedron Lett.* 37, 3429–3432.

- 50. Harre, M., Haufe, R., Nickisch, K., Weinig, P., Weinmann, H., Kinney, W.A., and Zhang, X. (1998) Some Reaction Safety Aspects of Ruthenium-Catalyzed Allylic Oxidations of Δ<sup>5</sup>-Steroids in the Pilot Plant, Org. Proc. Res. Dev. 2, 100–104.
- Salvador, J.A.R., Sáe Melo, M.L., and Campos Neves, A.S. (1997) Copper-Catalyzed Allylic Oxidation of Δ<sup>5</sup>-Steroids by *t*-Butyl Hydroperoxide, *Tetrahedron Lett.* 38, 119–122.
- Ishii, Y., Nakayama, K., Takeno, M., Sakaguchi, S., Iwahama, T., and Nishiyama, Y. (1995) Novel Catalysis by N-Hydroxyphthalimide in the Oxidation of Organic Substrates by Molecular Oxygen, J. Org. Chem. 60, 3934–3935.
- 53. Foricher, J., Fuerbringer, C., and Pfoertner, K. (1991) Process for the Catalytic Oxidation of Isoprenoids Having Allylic Groups, U.S. Patent 5,030,739.
- 54. Jones, S.R., Selinsky, B.S., Rao, M.N., Zhang, X., Kinney, W.A., and Tham, F.S. (1998) Efficient Route to 7α-(Benzoyloxy)-3-dioxolane cholestan-24(*R*)-ol, a Key Intermediate in the Synthesis of Squalamine, *J. Org. Chem.* 63, 3786–3789.
- 55. Shu, Y., Jones, S.R., Kinney, W.A., and Selinsky, B.S. (2002) The Synthesis of Spermine Analogs of the Shark Aminosterol Squalamine, *Steroids* 67, 291–304.
- 56. Arsenou, E.S., Koutsourea, A.I., Fousteris, M.A., and Nikolaropoulos, S.S. (2003) Optimization of the Allylic Oxidation in the Synthesis of 7-Keto- $\Delta^5$ -steroidal Substrates, *Steroids 68*, 407–414.
- Feldberg, L., and Sasson, Y. (1994) Copper-Catalyzed Oxidation of Hydroxy Compounds by *tert*-Butyl Hydroperoxide Under Phase-Transfer Conditions, J. Chem. Soc., Chem. Commun., 1807.
- 58. Parish, E.J., and Wei, T.Y. (1987) Allylic Oxidation of  $\Delta^5$ -Steroids with Pyridinium Chlorochromate (PCC) and Pyridinium Dichromate (PDC), *Synth. Commun.* 17, 1227–1233.
- 59. Fieser, L.F. (1953) Cholesterol and Companions. III. Cholestanol, Lathosterol, and Ketone 104, *J. Am. Chem. Soc.* 75, 4395–4403.
- Petrow, V.A., Rosenheim, O., and Starling, W.W. (1943) Acyl Migration in Steroids, J. Chem. Soc., 135–139.
- 61. Guillemonat, A. (1939) Oxidation of Ethylenic Hydrocarbons with Selenium Dioxide, *Ann. Chim. Appl.* 11, 143–211.
- 62. Hori, T., and Sharpless, K.B. (1979) Conversion of Allylic Phenylselenides to the Rearranged Allylic Chlorides by *N*-Chlorosuccinimide. Mechanism of Selenium-Catalyzed Allylic Chlorination of β-Pinene, *J. Org. Chem.* 44, 4208–4210.
- 63. Smith, M.B. (1994) Organic Synthesis, 1st edn., p. 156, Mc-Graw-Hill, New York.

[Received September 14, 2004; accepted September 15, 2004]