

**INDOLEALKYLAMINE METABOLISM: SYNTHESIS OF DEUTERATED
INDOLEALKYLAMINES AS METABOLIC PROBES**

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SUMMARY

The synthesis of the deuterium labeled, endogenously occurring, indolealkylamine hallucinogens *N,N*-dimethyltryptamine and 5-methoxy-*N,N*-dimethyltryptamine *via* reduction of amide intermediates with lithium aluminum deuteride is described. The compounds were characterized with ¹H, ²H and ¹³C NMR. These compounds were synthesized for use as probes for investigating the metabolism of these compounds by MAO *via* the *in vivo* kinetic isotope effect.

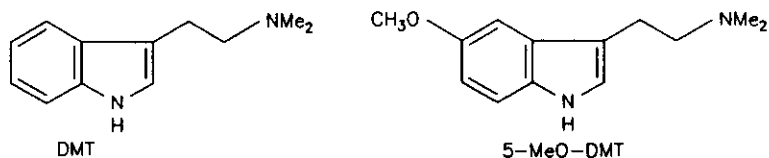
KEY WORDS: Deuterium Label, Endogenous Hallucinogen, Indolealkylamine, Monoamine Oxidase, Metabolism, *In Vivo* Kinetic Isotope Effect.

INTRODUCTION

The indolealkylamine hallucinogens *N,N*-dimethyltryptamine (DMT, 4) and 5-methoxy-*N,N*-dimethyltryptamine (5-MeO-DMT) have been identified as normal constituents of human blood,¹⁻⁶ urine,⁷⁻¹⁰ cerebrospinal fluid^{11,12} (CSF) as well as in a variety of plant species. DMT has also been identified as a putative neurotransmitter or neuromodulatory

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substance in rat brain.¹³ The indole-*N*-methyltransferase enzymes capable of synthesizing DMT and 5-MeO-DMT from tryptamine derived from L-tryptophan and S-adenosyl-methionine have been described and characterized in human lung, brain, blood and CSF and in various mammalian species.¹⁴

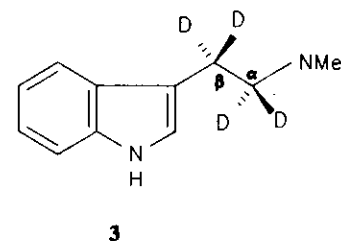


Numerous groups have attempted to relate mental disorders such as schizophrenia to high brain concentrations of these compounds resulting from perhaps a metabolic error, but a clear relationship between the two has not yet been delineated. However, the *in vivo* production of these interesting compounds strongly suggests that they serve some physiological role which is not yet understood.

For a number of years our group has been interested in studying the pharmacological properties of DMT and related indolealkylamines, in particular the *in vivo* metabolism. Indole-3-acetic acid (IAA) has been identified as the major *in vivo* metabolite of DMT. IAA is formed presumably *via* rapid oxidative deamination by monoamine oxidase (MAO) and is then excreted in urine.¹⁵ These studies are in good agreement with more recent studies which have shown that DMT is essentially cleared from whole rat brain in *ca.* 30 minutes and that DMT is undetectable in the liver and blood plasma after 30 minutes.¹⁶ The dominance of the deamination pathway makes it difficult to study the minor metabolites. Traditionally this difficulty has been surmounted by pre-treating animals with a MAO inhibitor such as pargyline. Inhibition of the major catabolic route leads to "shunting" to the minor metabolic routes, facilitating the study of the minor metabolites. Unfortunately pre-treatment of animals with pargyline can give experimental results which are difficult to interpret since pargyline also inhibits the *N*-oxidation and demethylation of

DMT by 90%.¹⁷ This observation suggests that the reported potentiation of the behavior-disrupting effects and the reported tissue levels of DMT measured in animals pre-treated with pargyline may not have been solely due to MAO inhibition.

Mechanistically, the deamination step presumably involves abstraction of an α -proton in the rate determining step of the reaction followed by deamination. This assumption led our group to speculate that substitution of the α and β protons of the ethylamine side-chain with deuteriums would produce the same effect *via* an *in vivo* kinetic isotope effect as pre-treatment with an appropriate MAO inhibitor (3).



Thus a ²H-labeled compound would be an attractive alternative to pre-treatment with a MAO inhibitor for metabolic studies.

The results of the metabolism study using $\alpha,\alpha,\beta,\beta$ -tetradeutero-*N,N*-dimethyltryptamine ($\alpha,\alpha,\beta,\beta$ -[²H]₄DMT, 3) have been published.¹⁸ The labeled compound was metabolized at a significantly slower rate than proteo-DMT (4) and has indeed proved useful as a metabolic probe for studying the minor metabolic products. Although we concluded that the α -deuteriums were responsible for the observed isotope effect it is impossible to measure the contribution of the α -deuteriums alone, since the ethylamine side-chain was fully deuterated. Although the β -deuteriums are not involved mechanistically, it has been demonstrated in a similar system that the β -deuteriums of aromatic ethylamines change the rate of deamination and produce a small rate enhancement. We are now interested in demonstrating unambiguously that the α -position is responsible for the

observed effect and is the only position involved in the deamination step. For this reason, our present study requires compounds labeled only in the α -position.

In this paper the synthesis and spectral properties of α,α -[^2H]₂-*N,N*-dimethyltryptamine (7) and α,α -[^2H]₂-5-methoxy-*N,N*-dimethyltryptamine (10) and a convenient synthesis of $\alpha,\alpha,\beta,\beta$ -[^2H]₄-*N,N*-dimethyltryptamine (3) is presented. Complete ^1H , ^2H and ^{13}C NMR assignments for $\alpha,\alpha,\beta,\beta$ -[^2H]₄-5-methoxy-*N,N*-dimethyltryptamine (11, isotopic purity of 97.5%) are also described.¹⁹

RESULTS AND DISCUSSION

Benington and Morin previously synthesized 3 in four steps from indole for use as an internal standard, however, its synthesis was not reported.²⁰ In their synthesis indole was acylated with oxalyl chloride to give the 3-substituted indole which was immediately reacted with ethereal dimethylamine to give the keto-amide (2). Reduction of the keto-amide with lithium aluminum deuteride (LAD) gave (3). Our group now utilizes commercially available indole-3-glyoxylic acid which affords the compound of interest in three steps. In a one-pot reaction the acid is converted to the acid chloride 1 with thionyl chloride which is not isolated but is immediately converted to the same keto-amide 2 by saturating the solution with dimethyl amine gas. In our hands, the preparation of 1 from the acid required low temperature and low concentration in order to prevent highly colored by-products. Attempts to isolate the acid chloride for analysis failed. The ^1H and ^{13}C spectra of 2 were rather complicated due to the formation of mesomeric forms and also due to the hindered rotation of the amide group. The ^{13}C spectrum of 2 has been reported but the resonances due to the mesomeric forms were not given.²¹ We have tabulated the non-mesomeric assignments in Table 2 and the mesomeric resonances are reported in the Experimental. Reduction of the keto-amide 2 with LAD gave 3 which was readily purified by sublimation under diminished pressure. For spectral comparisons, we also reduced a small amount of 2 with

LAH to give proteo-DMT (4). Complete ^1H NMR assignments are given in Table 1 and the ^{13}C assignments are described in Table 2. In the ^1H NMR of 3, the α and β -protons appear as apparent triplets centered at δ 2.95 and 2.64, respectively, and the ^{13}C spectrum

TABLE 1.

Compound	^1H NMR DATA (6) FOR DEUTERATED <i>N,N</i> -DIMETHYLTRYPTAMINE ANALOGS, INTERMEDIATES AND 5-METHOXY- <i>N,N</i> -DIMETHYLTRYPTAMINE ANALOGS AND INTERMEDIATES.									
	H-1	H-2 ($J_{1,2}$)	H-4' ($J_{4,3}$)	H-5' ($J_{5,6}$)	H-6 ($J_{6,7}$)	H-7	H- α ($J_{\text{H-4H}}$)	H- β	CH ₃	
2			7.84 (8.7)d			7.14				2.17
3	8.13	7.00 (2.1)d	7.61 (8.1)d	7.17 (7.8)vt	7.11 (6.9)vt	7.34				2.34
4	8.14	7.00 (1.5)d	7.69 (7.8)d	7.12 (7.8)vt	7.11 (6.9)vt	7.3	2.95 (8.1)vt	2.64 (8.1)vt		2.34
6	8.24	7.09 (0)bs	7.65 (7.8)d	7.20 (7.8)vt	7.13 (7.8)vt	7.40		3.83		3.03 2.98
7	8.22	7.02 (1.8)d	7.61 (7.8)d	7.18 (7.5)vt	7.11 (6.9)vt	7.35		2.34		2.17
9	8.20	7.04 (2.1)d	7.09 (2.1)d		6.88 (8.7)dd	7.23		3.79		3.03 2.98 3.88
10	7.88	7.00 (1.8)d	7.05 (1.8)d		6.86 (8.7)dd	7.25		2.90		2.34 3.86
11	7.04 (2.4)d	6.64 (3.6)d	7.40 (3.6)d		6.98 (9.0)d	8.32				2.34 3.86

¹ The values for H-4 and H-7 may be reversed.
² The values for H-5 and H-6 may be reversed.

showed two aliphatic resonances at δ 60.0 (α -carbon) and 23.0 (β -carbon). The remaining aromatic carbons were consistent with those previously reported. In the ^1H and ^{13}C NMR spectra of 3, there were no resonances at δ 2.95 and 2.64 and the resonances for the α and

β -carbons were absent. The remaining ^{13}C signals were assigned based on 4 and the ^2H NMR showed two singlets at δ 2.92 (α) and 2.62 (β) in good agreement with structure 3.

Reaction of indole-3-acetic acid with thionyl chloride gave the acid chloride 5 which was reacted with dimethylamine to give the amide 6. Reduction with LAD afforded 7 which was purified by sublimation. Characterization of 7 by ^1H NMR showed that the β -protons were shifted upfield to δ 2.34 and now appeared as a singlet. The ^{13}C spectrum lacked the signal at δ 60.0 and the β -C was shifted upfield 0.2 ppm to δ 22.8 consistent with adjacent ^2H . The ^2H spectrum showed one singlet at δ 2.61.

Using the same procedure, 5-methoxyindole-3-acetic acid was converted to the acid chloride 8 which was then converted to the amide 9. Usual reduction and purification afforded 10. The ^1H NMR showed a singlet at δ 2.90 and the ^{13}C NMR showed a greatly attenuated peak around δ 60.0. The ^2H NMR showed a singlet at δ 2.59. We also characterized 11 by NMR. The ^{13}C NMR lacked signals for the α and β -carbons and the ^2H NMR showed two singlets at δ 2.87 (α) and 2.59 (β).

EXPERIMENTAL

General Methods. All reagents and solvents were of the highest available purity and were used without further purification. Lithium aluminum deuteride (98%) was obtained from Aldrich Chemical Company. Thin layer chromatography (TLC) analyses were performed on Kieselgel aluminum backed silica gel 60 F₂₅₄ plates (0.2 mm) obtained from E. Merck and were visualized using an ultraviolet light (254 nm) or I₂. Gas chromatography-mass spectrometry was achieved on a Hewlett-Packard 5985 spectrometer operating at 70 eV. Isotopic purity measurements were made by mass spectrometry and calculations are based on comparisons of the spectra with the corresponding proteo-compounds. Melting points were recorded in capillary tubes on a Mel-Temp apparatus and are uncorrected. All ^1H NMR spectra were recorded at 300 MHz with a Nicolet Fourier

TABLE 2.

Compound	^{13}C NMR DATA (δ) FOR DEUTERATED N,N-DIMETHYLTRYPTAMINE ANALOGS, INTERMEDIATES AND 5-METHOXY-N,N-DIMETHYLTRYPTAMINE ANALOGS AND INTERMEDIATES.											CH ₃	OCH ₃	
	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C- α	C- β	CH ₃			
2	139.0	112.1	126.5 ^b	122.4 ^b	121.6 ^b	112.1	126.4	133.9					34.2	
3	122.0	111.1	119.2	119.2	121.9	112.8	127.5	136.3	a	a			45.5	
4	122.3	111.2	118.1	118.1	120.7	112.5	127.2	136.1	60.0	23.0			45.1	
6	123.4	111.2	118.7	120.9	118.2	108.1	127.2	136.0	170.5	37.2			34.9 34.6	45.0
7	122.3	111.2	118.0	120.7	118.0	112.5	127.2	136.1	a	22.8			31.3 37.8	55.8
9	123.5 ^c	112.0 ^c	100.3 ^c	153.8	112.0 ^c	108.3	127.4	131.1	171.7	35.6			45.4 55.9	55.9
10	122.3 ^c	111.9 ^c	100.6 ^c	153.8	111.9 ^c	113.9	127.7	131.4	a	23.5			45.5 55.9	55.9
11	122.2 ^c	111.8 ^c	100.7 ^c	153.8 ^c	112.0 ^c	114.0	127.8	131.4	a	a			45.5 55.9	55.9

a. This peak was greatly attenuated and was barely discernable from the baseline.

b. These values are from Reference 20.

c. These values may be interchanged.

Transform Spectrometer in CDCl_3 or DMSO-d_6 . Resonances are reported downfield from internal tetramethylsilane. Multiplicities are reported as follows: s, singlet; d, doublet; ∇ t, an apparent triplet, i.e. $J_{a,b} = J_{b,c}$; m, multiplet, b, broad. ^{13}C NMR spectra were recorded at 75.5 MHz. ^2H NMR spectra were recorded at 46.07 MHz using a one pulse experiment in CHCl_3 with 1-2% CDCl_3 serving as an internal standard. The ^{13}C NMR spectra were recorded at 75.5 MHz. Infrared spectra were recorded on a Nicolet IR\42 spectrometer. Microanalyses were performed by Atlantic Microlabs, P.O.Box 2288, Norcross, Ga. 30091-9990.

2-(3-Indolyl)-glyoxal Chloride (1). A solution of indole-3-glyoxylic acid (1.0 g, 5.3 mmol) in ether (100 mL) was stirred and cooled in a dry-ice bath for 15 min. SOCl_2 (2.0 mL, 17 mmol) was then gradually added to the solution. TLC analysis (10% MeOH-90% Tol) of the mixture showed complete disappearance of indole-3-glyoxylic acid after 1 h and formation of a higher running compound with R_f 0.39. The product was diluted with a large amount of dry ether and used directly without purification.

2-(3-Indolyl)-N,N-dimethylglyoxalamide (2). Dimethylamine gas was passed through the etherial solution of **1** for 3-5 min and the reaction mixture was stirred continuously for an additional 20-30 min. Excess solvent was removed to give **2** (0.89 g, 4.1 mmol) as a solid in 77% yield based on indole-3-glyoxylic acid. $\text{Mp} = 184-185^\circ\text{C}$ (lit. $184-185^\circ\text{C}$)¹⁹; ^{13}C NMR δ (CDCl_3 - DMSO-d_6) 136.8, 136.5, 135.9, 126.4, 125.3, 124.0, 123.8, 123.2, 122.8, 122.2, 121.8, 112.64, 112.0, 40.2, 39.9, 39.6, 39.4, 37.9, 37.4, 35.1, 34.2; MS 216 [M+], 144 (100%), 116, 89, 72; IR 1618 cm^{-1} (CO).

$\alpha,\alpha,\beta,\beta$ -[^2H]₄-N,N-Dimethyltryptamine (3). To a stirred suspension of LAD (0.1 g, 2.4 mmol, 98%) in dry ether (8 mL) was gradually added the amide **2** (0.1 g, 0.46 mmol) in CH_2Cl_2 (5 mL). The mixture was refluxed for 3-4 h in an oil bath, cooled in an ice bath, and treated with several drops of water to decompose excess LAD reagent. The reaction

was vacuum filtered to remove any remaining solids, dried (MgSO_4), and solvents removed. The yield was 67% (0.06 g, 0.31 mmol). $\text{Mp} = 47-49^\circ\text{C}$; MS 192 [M+], 132, 60 (100%). Isotopic purity = 94%.

N,N-Dimethyltryptamine (4). LAH reduction of **2** gave **4** in 76% yield (0.066 g, 0.35 mmol). $\text{Mp} = 44-46^\circ\text{C}$ (lit. $45-46^\circ\text{C}$); MS 188 [M+], 130, 77, 58 (100%).

2-(3-Indolyl)-acetyl Chloride (5). Indole-3-acetic acid (2.0 g, 11.4 mmol) was converted to the acid chloride (ether, 200 mL; SOCl_2 , 2.0 mL, 17 mmol) using the procedure described for the synthesis of **1**.

2-(3-Indolyl)-N,N-dimethylacetamide (6). Acid chloride **5** was converted to the amide **6** using dimethylamine as described for the synthesis of **2**. Sublimation under diminished pressure gave 1.6 g (7.9 mmol, 69%) based on indole-3-acetic acid. $\text{Mp} = 117-119^\circ\text{C}$. MS 202 [M+], 130 (100%), 77, 72; IR 1634 cm^{-1} (CO).

Anal. Calcd for $\text{C}_{12}\text{H}_{14}\text{N}_2\text{O}$: C, 71.26; H, 6.98; N, 13.85. Found C, 71.13; H, 6.96; N, 13.75.

α,α -[^2H]₂-N,N-Dimethyltryptamine (7). LAD (0.1 g, 2.4 mmol, 98%) was suspended in dry ether (8 mL). The mixture was stirred and the amide **6** (0.25 g, 1.24 mmol) in CH_2Cl_2 (100 mL) was added over 5 min. The reaction was refluxed for 2-3 h in an oil bath at which time TLC analysis (MeOH) indicated disappearance of **6** and formation of a new spot at the origin. Workup as described for the synthesis of **3** and sublimation gave 0.16 g (0.84 mmol) of **7** in 68% yield. $\text{Mp} = 44-46^\circ\text{C}$ (lit. $\text{Mp} = 44-46^\circ\text{C}$)¹⁹. MS 190 [M+], 130, 60 (100%). Isotopic purity = 97%.

Anal. Calcd for $\text{C}_{12}\text{H}_{14}\text{D}_2\text{N}_2$: C, 75.76; H plus D as H, 9.52; N, 14.73. Found C, 75.09; H plus D as H, 8.56; N, 14.45.

2-(5-Methoxy-3-indolyl)-acetyl Chloride (8). 5-Methoxyindole-3-acetic acid (0.5 g, 2.44 mmol) was converted to the acid chloride (CH_2Cl_2 , 100 mL; SOCl_2 , 1.0 mL, 8.5 mmol) using the procedure described for the synthesis of 1.

2-(5-Methoxy-3-indolyl)-N,N-dimethylacetamide (9). The acid chloride 8 was diluted with CH_2Cl_2 and immediately treated with dimethylamine gas. The excess solvent was removed and the crude product sublimed under diminished pressure. The yield was 74% (0.42 g, 1.8 mmol) based on 5-methoxyindole-3-acetic acid. $\text{Mp} = 78\text{--}80^\circ\text{C}$. $\text{MS } 232 \text{ [M+]}$, 160 (100%), 145, 117, 72; $\text{IR } 1629 \text{ cm}^{-1} \text{ (CO)}$.

Anal. Calcd for $\text{C}_{13}\text{H}_{16}\text{N}_2\text{O}_2$: C, 67.22; H, 6.94; N, 12.06. Found C, 67.00; H, 6.97; N, 12.00.

$\alpha,\alpha\text{-}[^2\text{H}]_2\text{-5-Methoxy-N,N-dimethyltryptamine (10)}$. A suspension of LAD (0.1 g, 2.4 mmol, 98%) in dry ether (8 mL) was stirred and the amide 9 (0.25 g, 1.08 mmol) in CH_2Cl_2 (10 mL) gradually added. The mixture was refluxed for 2-3 h in an oil bath and then cooled to room temperature. Usual workup and purification gave 10 in 68% yield (0.16 g, 0.73 mmol) based on the amide 9. $\text{Mp} = 49\text{--}51^\circ\text{C}$. $\text{MS } 220 \text{ [M+]}$, 176, 160, 145, 132, 117, 60 (100%). Isotopic purity = 99.7%.

Anal. Calcd for $\text{C}_{13}\text{H}_{16}\text{D}_2\text{N}_2\text{O}$: C, 70.88; H plus D as H, 9.14; N, 12.72. Found C, 70.22; H plus D as H, 8.32; N, 12.09. Despite repeated sublimations and drying, we were unable to resolve the elemental analysis difference.

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